

Research Article

Assessment of Seed Priming on Germination Improvement of *Lathyrus sativus* under Allelopathic Components of *Juglans regia*

Mojtaba Akhavan Armaki

Department of Agriculture

Payame Noor University (PNU), Karaj, Iran

Email: mtakhavan@ut.ac.ir

ABSTRACT

The objective of this study was to evaluate the effectiveness of seed priming in improving seed germination and seedling vigor of *Lathyrus sativus* under laboratory conditions. Growth stimulators included: gibberellic acid (125,250 and 500 ppm) and salicylic acid (100,200 and 300 mg/lit). Extract of *Juglans regia* as allelopathic compounds consist of 0, 25, 50, 75 and 100 percentages. This experiment was carried out as factorial experiment based on a randomized completely design with four replications. The results showed that *Juglans regia* extract had inhibitive effect on germination and early seedling growth of *Lathyrus sativus*. Early seedling growth of *Lathyrus sativus* increased by pretreatment of seeds in growth stimulators so that the highest effect was observed in 250 ppm level of gibberellic acid. The growth stimulators don't have any effect on germination speed. Interaction effects of allelopathic and pretreatment with chemical stimulators were significance on germination percentage, root, shoot and plant length and seed vigour index.

Keywords: Germination, Allelopathy, *Juglans regia*, *Lathyrus sativus*

INTRODUCTION

Germination is one of the most important stages on life cycle of plant since it controls establishment of plants and determines final function of them. Poor establishment of plant is a main reason of decreasing its function in arid and semiarid environment [1-10]. In this areas, soil environment often are not suitable for high germination and growth of plants. Biotic and abiotic factors such as lack or extras of water and high elements can decrease the germination speed and percentage or inhibit them completely [2]. Improvement of germination can increase establishment of plant spatially in stress condition [11]. Allelopathy, as a stress, is defined as direct and indirect effects of allelochemical compounds resulted from organism which may have inhibitive or stimulative effects on the same or different

organism. Allelopathic components restrict growth of plant through interaction in important physiological process such as change in cell wall structure, infiltration and function of membrane, prevention of cell division and activity of some enzymes. These components also can cause effect on equilibrium of plant hormones, absorption of nutrient elements, displacement of stomata, photosynthesis, respiration, protein synthesis and pigment and change in DNA and RNA structures [8]. This phenomenon has direct effect on plants in agricultural ecosystems or indirect effects in biological or non biological process through the same or the other plants [14]. Priming is used to improve germination, reduce germination time and embryo emergency, and improve establishment and performance [9-20-24]. Priming

also applies in increasing of seed vigourity and reducing of losses from late plantation. Many researchers have reported that priming can increase germination percentage and emergence of weakened or damaged seeds [12].

Different physiological and biochemical effects from salicylic acid on plant systems have been observed that include ion absorption, membrane permeability, mitochondrial respiration, and effect on stomata, growth rate and photosynthesis rate [21]. Furthermore, salicylic acid causes production of phenolic matter which acts as blockage in cell wall and then decreases losses of water and also inhibits spread of diseases. Phenols acts as antioxidant in plant and causes to trap free radicals by antioxidant process [5]. It is determined that salicylic acid decreased ion leakage and accumulation of toxic ions significantly in plants [17-25] and caused decrease the effect of environmental stress via increase of hormones such as auxins and cytokinins [22]. Saberi *et al.* (2011) reported that pretreatment the seeds of *Agropyron elongatum* and *Bromus inermis* by growth stimulators (gibberellic acid, potassium nitrate and salicylic acid) decreased the allelopathic effect of *Thymus kotschyanus*. It has been pointed to salicylic acid as intermediate for reacting to abiotic stress [4-18]. Pre-treatment with chemical substance had been known as a simple technique, low cost and risk to improve germination and seedling. The objective of this study was to evaluate the effectiveness of seed priming in improving seed germination and seedling vigour of *Lathyrus sativus*, in response to allelopathic effects of *Juglans regia* under laboratory conditions. *Juglans regia* is one of the most important plants used to prevent soil erosion and to recover the plant cover in studied area. This plant also used in farmland as windbreak and medical plant. In studied area which consists of 6000 hec, many plants (range species, cultivatable and medical plant) were cultivated based on different goals. This research was conducted to increase the resistance of *Lathyrus sativus* in facing inhibitory effect of *Juglans regia* by using

growth stimulators (include: gibberellic acid and salicylic acid as pre-treatment).

MATERIAL AND METHODS

This research was conducted to determine the effect of growth stimulators used to reduction of allelopathic effect of *Juglans regia* on germination and primary growth of *Lathyrus sativus*. To meet this aim, at first, aerial and underground parts of *Juglans regia* were collected from Chah Nime, Zabol, Iran. After air drying at room temperature, 5 g of powder was picked and mixed in 100 mL water, placed on a shaker for 24 h then centrifuged at 3000g for 15 min. The obtained mixture was filtered using Whatman 1 filter paper. Concentrations of 25, 50, 75 and 100% were prepared using centrifuged solution. Seeds of *Lathyrus sativus* (collected from Chah Nime's farm and rangeland) were disinfected by using 5% solution of sodium hypochlorite before starting of test and were washed by using distilled water several times. Then seeds were pretreated using salicylic acid 100, 200 and 300 mg for 10 hours and using gibberellic acid 125, 250 and 500 ppm for 24 hours at 25°C temperature and distilled water were used as control treatment simultaneously. All seeds were washed with distilled water after soaking period and then were placed into petri dishes with dimensions of 9cm on a filter paper (Watman 1) after being dried in order to test different stress conditions with various concentration of allelopathic extract related *Juglans regia*. Petri dishes were sterilized for 48 hours in the oven at 20°C before placing seeds. Germination test was performed using factorial test (5×7) in completely randomized design with 4 replications (25 seeds per Petri dishes) in different concentration of extract related to *Juglans regia* (0, 5, 25, 50 and 75 percentage) and 25°C in the germinator. Germinated seeds that had length more than 2mm were counted each day over a period of 10 days [16] and germination percentage, germination speed, root length shoot length, plant length and vigour index of seed were measured. Germination percentage (Camberato

and Mccarty, 1999) and germination speed were measured based on equations at follow:

(1) Germination percentage

$$GP = \frac{\sum G}{N} \times 100$$

GP: germination percentage, G: number of germinated seeds, N: number of seeds

(2) germination speed

$$GR = \frac{\sum_{i=1}^n S_i}{D_i}$$

S_i : number of germinated seed at each counting,
D_i: number of day until n counting, n: numbers of counting

(3) plant length = root length + shoot length

(4) vigour index

$$V_i = \frac{\% Gr \times MSH}{100}$$

V_i : vigour index, MSH: mean of plant length (root length + shoot length) per mm, Gr: Germination percentage

The obtained data was analyzed using analysis of variance (ANOVA). Means were compared at the 5% level of significance using Duncan's multiple range tests with statistical software MSTAT-C version 2.00.

RESULTS

Results of variance analysis (table 1) showed that growth stimulators and various concentrations of *Juglans regia* had significant effect on all studding properties of *Lathyrus sativus* species (P<0.01). Also interaction of chemical stimulators and various concentrations of extract had significant effect studding properties except for shoot length (P<0.05).

| Properties | | A | B | A* B | Error |
|------------------------|----|-----------|------------|----------|-----------|
| Germination percentage | df | 6 | 4 | 24 | 105 |
| | ss | 14084.3 | 10707.1 | 2922.9 | 7825.0 |
| | ms | 2347.4 | 2676.8 | 121.8 | 74.5 |
| | F | 31.5** | 35.9** | 1.6* | - |
| Germination speed | df | 6 | 4 | 24 | 105 |
| | ss | 171.9 | 117.7 | 15.1 | 21.2 |
| | ms | 28.6 | 29.4 | 0.6 | 0.2 |
| | F | 141.8** | 145.6** | 3.1** | - |
| Root length | df | 6 | 4 | 24 | 105 |
| | ss | 403.4 | 538.1 | 45.9 | 33.4 |
| | ms | 67.2 | 134.5 | 1.9 | 0.3 |
| | F | 210.7** | 421.7** | 5.9** | - |
| Shoot length | df | 6 | 4 | 24 | 105 |
| | ss | 165 | 196.4 | 12.8 | 31.3 |
| | ms | 27.5 | 49.1 | 0.5 | 0.2 |
| | F | 92** | 164.3** | 1.7* | - |
| Plant length | df | 6 | 4 | 24 | 105 |
| | ss | 1056 | 1381.1 | 58.4 | 75.5 |
| | ms | 176 | 345.2 | 2.4 | 0.7 |
| | F | 244.6** | 479.9** | 3.3** | - |
| Seed vigor | df | 6 | 4 | 24 | 105 |
| | ss | 9535540.9 | 14412756.5 | 819220.8 | 1199286.8 |
| | ms | 1589256.8 | 3603189.1 | 34134.2 | 11421.8 |
| | F | 139.1** | 315.5** | 3.0** | - |

** : significant differences between treatments at 1% level; ns: non-significant differences between treatments; A = pretreatment; B = allelopathic extract.

Germination percentage and speed

The results of this research showed that germination percentage of *Lathyrus*

sativus decrease by increasing various concentrations of allelopathic extract of *Juglans regia*. Differences were significance between

control treatment and various concentrations of extract. All the growth stimulators could increase germination percentage of *Lathyrus sativus* seeds comparing to control treatment (fig. 1) so that maximum of germination percentage was related to various concentrations of gibberelic acid. Interaction effects of growth stimulators and various concentrations of *Juglans regia* on germination percentage of *Lathyrus sativus* seeds

were significant ($P < 0.01$) (Fig. 1). Results showed that germination speed of seeds that were exposure of various concentrations of extracts had significant differences with the control treatment. Using of the growth stimulators doesn't have any effect on germination speed so that they decreased germination speeds compare to control treatment (fig. 2).

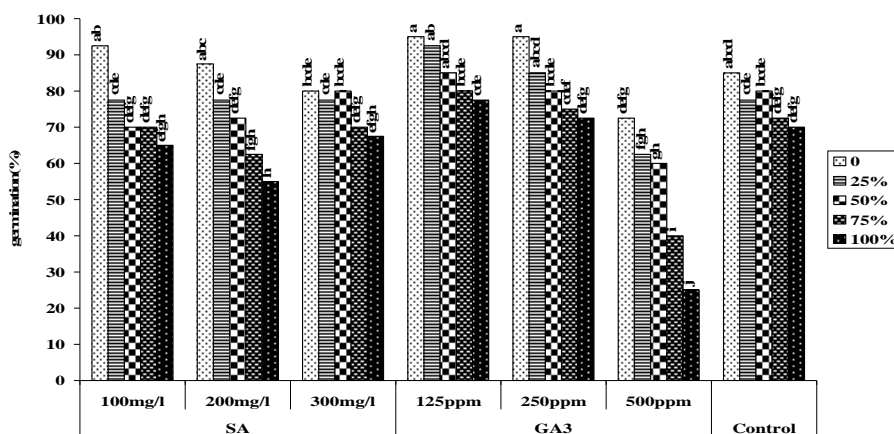


Fig. 1. Interaction comparison of growth stimulators and various concentrations on germination of *Lathyrus sativus*

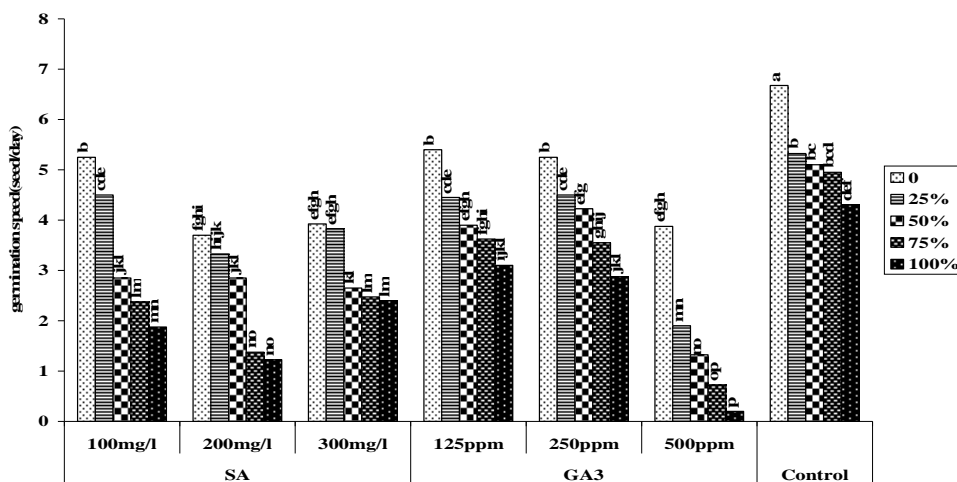


Fig. 2. Interaction comparisons of growth stimulators and various concentrations on germination speed of *Lathyrus sativus*

Root, shoot and plant length

Interaction effect of growth stimulators and various concentrations of extract of *Juglans regia* were significant on root length. All stimulators improved root length in stress condition with extract of *Juglans regia* so that the highest root length was related to use various concentration of gibberellic acid (fig. 3).

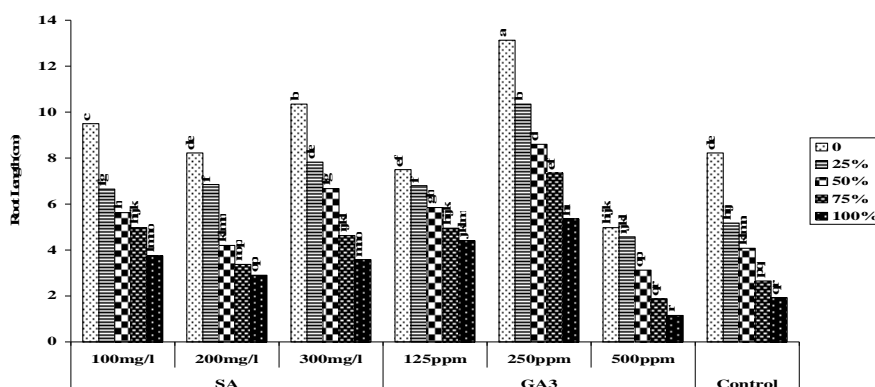


Fig. 3. Interaction comparison of growth stimulators and various concentrations on root length of *Lathyrus sativus*.

Results also indicated that Interaction effects of growth stimulators and various concentrations of extract of *Juglans regia* were significant on shoot length. The highest shoot length of *Lathyrus sativus* was related to use of gibberellic acid treatment in stress and non stress conditions. Various concentrations of *Juglans regia* reduced shoot length of the species. Growth stimulators caused an increase in shoot length in stress condition which differences were significant (Fig. 4).

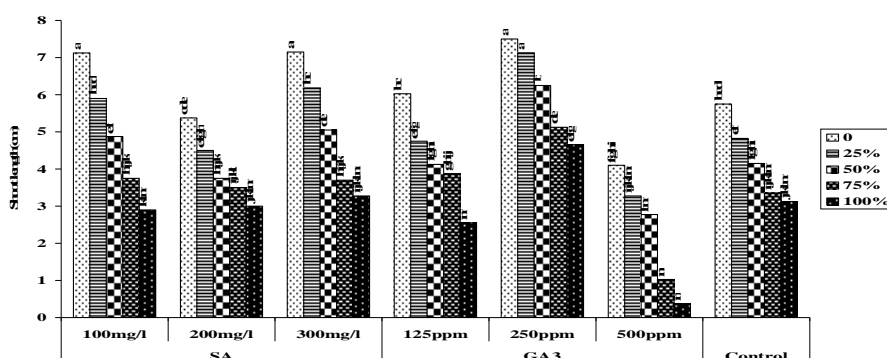


Fig. 4. Interaction comparison of growth stimulators and various concentrations on shoot length of *Lathyrus sativus*.

Mean comparison of data showed that interaction effect of growth stimulators and various concentrations of *Juglans regia* were significant on plant length so that plant length reduced by increasing concentration of *Juglans regia*. In reverse all concentrations of growth stimulators caused an increase in plant length of *Lathyrus sativus* in stress condition except for 500 ppm gibberellic acid. The highest its plant length was related to use of gibberellic acid at 250 ppm in stress and non stress conditions (fig. 5).

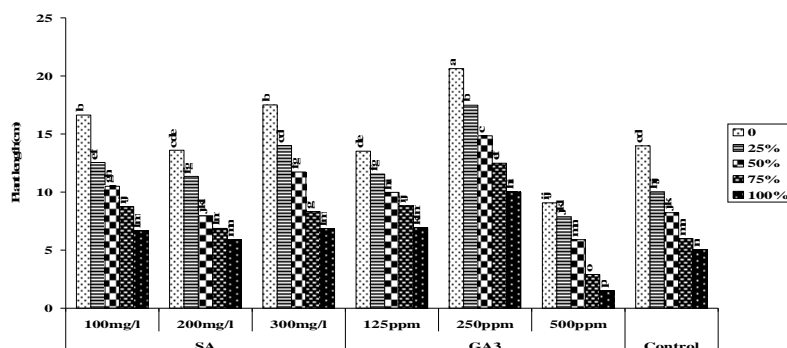


Fig. 5. Interaction comparison of growth stimulators and various concentrations on plant length of *Lathyrus sativus*.

Seed vigour index

Mean comparison showed that interaction effect of growth stimulators and various concentrations *Juglans regia* were significant on vigour index. Results showed that vigour index reduced by increasing the concentration of *Juglans regia* and it was significant by comparison to the control treatment. In reverse growth stimulators increased vigour of seeds as the highest increase was related to gibberellic acid at 250 ppm (fig. 6).

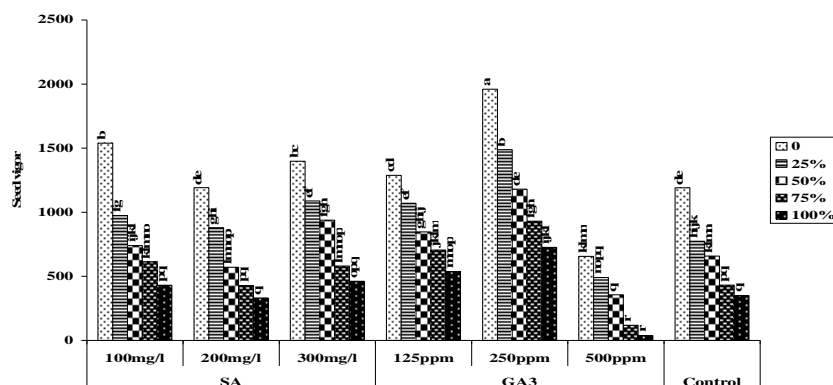


Fig. 6. Interaction comparison of growth stimulators and various concentrations on seed vigor of *Lathyrus sativus*.

DISCUSSION AND CONCLUSION

The objective of this research was to test effects of pretreatment (salicylic and gibberellic acid) on germination properties of *Lathyrus sativus* under stress condition. Results of this research showed that growth stimulators used in this study play a key role in reduction of stress induced by allelopathic compounds of *Juglans regia*. These results coincide with the results of Saberi *et al.* (2011) who stated growth stimulators such as gibberellic and salicylic acid could be used as stimulators for improving germination under allelopathic condition. Obtained results from Kang and Saltveit (2002) and Tasgin *et al.* (2003) also verify this hypothesis that salicylic acid is a suitable stimulator for seed germination. Salicylic acid increases germination by neutralizing free radicals or active oxygen (Hus and Sung, 1997), increasing of antioxidant such as ascorbate [3], decreasing of ion transfers and accumulation of toxic ions [17] and increasing of some plant hormones such as auxins and cytokinins [22]. In addition to the effect of salicylic acid in increasing plant growth in stress condition, this research confirms the importance of these phenolic compounds on improvement of the initial growth stage when seeds are exposed to stress conditions with *Juglans*

regia. Germination and early seedling growth decreased by increasing extract concentration. This may be because of the inhibitive effects of allelochemicals on gibberellic acid. Pretreatment with gibberellic acid increased germination and early seedling growth of *Lathyrus sativus* significantly under stress and non-stress conditions. This hormone has an important role in germination of seeds [19]. Gibberellic acid increases enzymatic synthesis such as hydrolytic enzymes. Synthesized enzymes transfer to the endosperm and cause digestion of reserve food and provide supply of energy for germination and growth. Delay or stimulation in digestion of reserve food may cause lack of production of respiration and consequently caused lack of ATP in seeds exposed to allelochemicals. Disorder in respiration results in limits in metabolic energy and causes decrease in germination and early seedling growth [7].

Overall, results indicated that pretreatment of seeds by gibberellic and salicylic acid causes improvement in germination properties of *Lathyrus sativus* under stress and non-stress conditions. It is suggested that 250 ppm of gibberellic acid was used as pretreatment to decrease the negative effects of *Juglans regia* on

Lathyrus sativus. Hence germination percentage and establishment of *Lathyrus sativus* must be increased by pretreatment with gibberellic acid before planting in field.

REFERENCES

1. Afzal I., (2005). Ph.D. Thesis, Agricultural University of Faisalabad, Pakistan, 266 p.
2. Ashraf M., M.R. Foolad, (2005). *Advances in Agronomy*, 88:223- 265.
3. Baalbaki R.Z., R.A. Zurayk, M.M. Blek, S.N. Tahouk, (1999). *Seed Sciences and Technology*, 27:291-302.
4. Bor M., F. Ozdemir, I. Turkan, (2003). *Plant Science*, 164:77-84.
5. Burguieres E., P. McCu, Y.I. Kwon, K. Shetty, (2007). *Bioresource Technology*, 98:1393-1404.
6. Camberato J., B. Mccarty, (1999). *South Carolina Turfgrass Foundation News*, 6: 68.
7. Cirac C., A.K. Ayan, K. Kevseroglu, (2004). *Pak. J. Biol. Sci.*, 7: 182-186.
8. Glass A.D.M., (1974). *J. Exp. Bot.*, 25:1104-1113.
9. Ghobadi M, M Shafiei-Abnavi, S Jalali-Honarmand, ME Ghobadi, GR Mohammadi, (2012). *Annals of Biological Research*, 3 (7):3156-3160
10. Harris D., A.K. Pathan, P. Gothkar, A. Joshi, W. Chivasa, P. Nyamudeza, (2001). *Agric. Syst*, 69: 151-164.
11. He Y.L., Y.L. Liu, Q. Chen, A.H. Bian, (2002). *Journal of Plant Physiology, Molecular and Biology*, 28: 89-95.
12. Horii A, P. McCue, K. Shetty, (2007). *Bioresource Technology*, 98: 623–632.
13. Hus J.L., J.M. Sung, (1997). *Physiologia Plantarum*, , vol. 100: 967-974.
14. Inderjit W.J., (2001). *Agronomy Journal*, vol. 93: 79-84.
15. Kang H.M., M.E. (2002). *Saltveit, Plantarum*, 115: 571-576.
16. Kaya M.D., G. Okçu, M. Atak Y. Çıkılı, Ö. Kolsarıcı, Europ. (2006). *J. Agronomy*, 24: 291–295.
17. Krantev A., R. Yordanova, T. Janda, G. Szalai, L. Popova, (2008). *J. Plant Physiol.*, 165: 920-931.
18. Naser-alavi S.M., G.h. Safari, M. Govahi, (2008). *The First National Iranian Seed Science and Technology*, Iran.
19. Ritchie S., S. Gilroy, (1998). *New Phytologist*, 140: 363–383.
20. Saberi M., A.R. Shahriari, F. Tarnian, M. Jafari, H. Safari, (2011). *Frontiers of Agriculture in China*, 5: 310-321.
21. Senaratna T., (2003). *Plant Growth Regulation*, vol. 30: 157-161.
22. Sharikova F., A. Sakhabutdinova, M. Bezrukova, R. Fatkhutdinova, D. Fatkhudinova, (2003). *Plant Sci.*, vol. 164: 317-322.
23. Tasgin E., O. Atic, B. Nalbantoglu, (2003). *Plant Growth Regul*, 41:231-236.
24. Zahedi SM, M Azizi, H Gheysari, (2012). *Annals of Biological Research*, 3 (8):4192-4194
25. Zhou Zh.Sh., K. Guo, A. Abdou Elbaz, Zh.M. Yang, (2009). *Environmental and Experimental Botany*, 65: 27-34.