

Research Article**Evaluation of Oxidative Stress Indices after Exposure to Malathion and Protective Effects of Vitamin C in Ovarian Tissue of Adult Female Rats.**Somayyeh Abbasabad Arab¹, Mohammad Reza Nikravesh*¹,Mahdi Jalali¹ and Ali Reza Fazel²¹Department of Anatomy and Cell Biology,
Mashhad University of Medical Sciences, Mashhad, Iran.²Microanatomy research center,
Mashhad University of medical Sciences, Mashhad, Iran.

*Corresponding Author: nikraveshmr@mums.ac.ir

ABSTRACT:

Background: Malathion is an organophosphate pesticide which is known to induce oxidative stress in the target tissues such as the reproductive system. The aim of present study was to evaluate the effects of malathion on malondialdehyde (MDA) and glutathione (GSH) levels in female rat reproductive tissue (ovary) and to assess the protective role of vitamin C.

Materials and Methods: In this experimental study, 30 adult female Wistar rats (weight range: 200-250 g) were divided into five groups, consisting of six rats: control group (no interventions), sham group (normal saline), experimental group 1 (malathion+normal saline 50 mg/kg), experimental group 2 (200 mg/kg of vitamin C), and experimental group 3 (similar doses of vitamin C and malathion). The animals were sacrificed after two weeks and MDA as a marker of lipid peroxidation and GSH content were measured in ovarian tissue.

Results: Malathion reduced GSH content and increased MDA level in ovary compared with the control group ($P < 0.001$). Vitamin C plus malathion increased GSH content but decreased malathion induced MDA elevation in rat ovarian tissue.

Conclusion: Oxidative stress contributes to DZN-induced ovarian toxicity. Our results concluded that vitamin E may have a protective role in this toxicity.

Keywords: Malathion, Glutathione, Malondialdehyde, Vitamin C, Ovary, Rat,

INTRODUCTION

Considering the widespread use of organophosphorus compounds in agriculture, industry, and veterinary medicine, studies on these compounds and their mechanisms of action are still under way (1, 2). So far, a wide range of effects have been attributed to organophosphorus compounds. The major mechanism of action in these compounds involves the inhibition of acetylcholinesterase and development of cholinergic crisis. However, many effects of these compounds are not associated with the inhibition of acetylcholinesterase (2). In Iran, organophosphorus compounds are considered as the third cause of toxicity and the main cause

of toxicity-induced mortality (3). One of the mechanisms involved in pesticide effects is the production of free radicals, followed by changes in the antioxidant system and lipid peroxidation of cell membranes. Under normal conditions, there is a balance between the production and elimination of free radicals. Imbalance in this process leads to oxidative stress and multiple pathological changes in cellular macromolecules such as DNA, lipids and enzymes. (4). Oxidative stress leads to the production of ROS and causes changes in enzymatic activities and antioxidant defense mechanisms in the body (5, 6). However, excessive production of ROS can damage the cells through lipid peroxidation

(7-10). Malondialdehyde (MDA) is a product of lipid peroxidation. MDA is in fact one of the important products resulting from the oxidation of polyunsaturated fatty acids. MDA measurement is one of the important indices of lipid peroxidation (11, 12). Thiol groups are reducing agents, existing at a concentration around 5 mM in animal cells (13). Thiol groups are sensitive to oxidation, and lipid peroxidation often results in a decline in their level (14). Glutathione belongs to thiol groups and plays an essential role in protecting body cells against damages such as oxidative stress (15). Antioxidants are a family of diverse compounds, which inhibit the excessive production of toxic free radicals and the induced damages (16, 17). Vitamin C as a vital antioxidant can change lipid oxidation parameters through reducing the production of oxidative stress (18). The purpose of this study was to evaluate the level of glutathione and MDA in ovarian tissues of malathion-induced female rats and to assess the protective effects of vitamin C.

MATERIALS AND METHODS

In this experimental study, 30 adult female Wistar rats (aged two months), with a weight range of 200-250 g, were randomly divided into five groups, consisting of six rats: experimental group 1 (daily administration of 50 mg/kg of malathion in normal saline solution for two weeks), experimental group 2 (daily administration of 200 mg/kg of vitamin C), experimental group 3 (daily administration of 50 mg/kg of malathion and 200 mg/kg of vitamin C), control group (no interventions), and sham group (administration of 50 mg/kg of normal saline for two weeks). Normal saline solution was used in this study, and malathion and vitamin C were intraperitoneally injected. After two weeks, the rats were sacrificed and MDA level was measured as an index of lipid peroxidation; also, the amount of glutathione in the ovaries of rats was estimated.

Chemicals

For this study, technical malathion was used. Malathion was dissolved in NaCl 0.9%. Malondialdehyde tetrabutylammonium, reduced GSH, DTNB [5,5 dithiobis-(2-nitrobenzoic

acid)] and vitamin C were purchased from Sigma company.

Lipid peroxidation test

The amount of lipid peroxidation was assessed through the measurement of MDA levels in ovary tissue of rats. MDA reacts with ThioBarbituric Acid (TBA) and produces a pink colored complex which has the maximum absorbance at 532 nm. Initially, 1 ml TBA (0.6%) and 3 ml phosphoric acid (1%) were added to homogenate 10% tissue in KCl, then the compound was heated for 45 min in a boiling water bath. After cooling the compound, 4 ml of n-butanol was added to it and then vortex-mixed was used for 1 min followed by centrifugation at 3000 g for 10 min. After this, the organic layers were removed and transferred to other tubes and absorbance level was read at 532 nm (40). A calibration curve was designed using MDA tetrabutylammonium. MDA levels were expressed by nmol/g tissue.

Reduced GSH test

GSH level measured in ovary by the method of Moron et al (41). The base of the work was the formation of yellow color after adding DTNB [5,5 dithiobis-(2-nitrobenzoic acid)] to compounds containing sulfhydryl groups. Therefore, 300 µl of homogenates tissue were blended with 300 µl of 10% Tricolor Acetic Acid (TCA) and vortexed. After centrifugation at 2500 g for 10 min, the upper layers were removed and mixed with reaction mixtures containing 2 ml phosphate buffer (pH:8) and 500 µl DTNB. After 10 min, the absorbance was read at 412 nm using a spectrophotometer (Jenway 6105 uv/vis, UK). At the end, the amount of GSH was determined based on a standard curve drawn with commercially available GSH (Sigma). The GSH Levels were expressed by nmol/g tissue

Statistical analysis

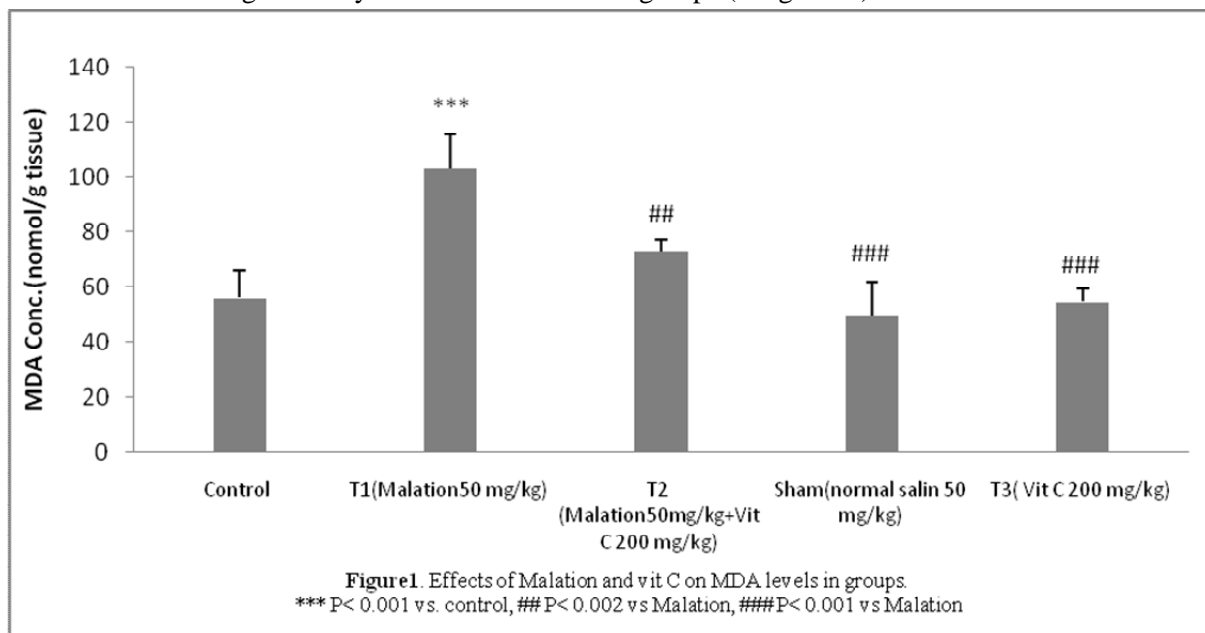
Results are expressed as mean ± SD. Statistical analysis was performed with ANOVA followed by Tukey-Kramer test to compare the differences between means. Differences were considered statistically significant at $P < 0.05$.

RESULTS

The effect of vitamin C on lipid peroxidation of malathion-treated rats

The results of the present study showed a significant increase in the level of MDA in experimental group 1 (malathion-induced), compared to the control group. In addition, the level of MDA significantly decreased in

experimental group 3 (malathion+vitaminC), compared to experimental group 1. However, the MDA level was not significantly different between the control, sham, and vitamin C groups (Diagram 1).



The effect of vitamin C on the level of glutathione in ovarian tissues of rats following the injection of malathion

In the present study, malathion-treated rats experienced a significant decline in the level of glutathione in experimental group 1, compared to the control group. Also, the level of glutathione significantly increased in experimental group 3 (vitamin C+malathion), compared to experimental group 1. Also, the control, sham, and vitamin C groups were not significantly different in terms of glutathione level (Diagram 2).

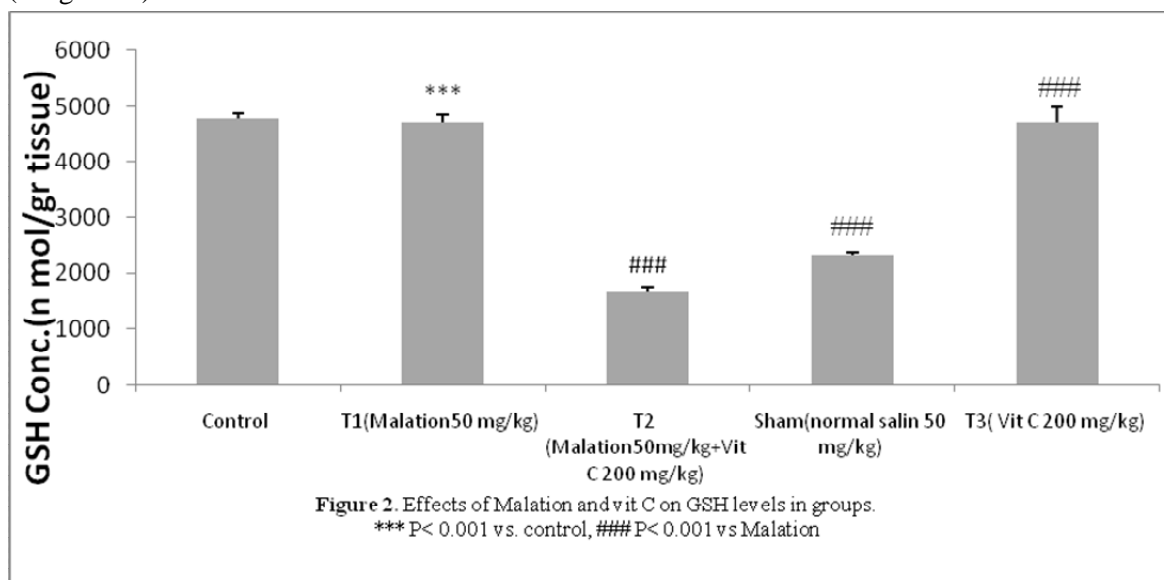


Table 1. Mean of GSH and MDA among various groups

Treatments	Glutathione	Malondialdehyde
Control	4783.861±93.23	56.04±10.07
Sham(normal saline 50 mg/kg)	4711.593±150.45	49.91±12.09
T1(Malation 50 mg/kg)	1665.815±89.43	103.13±12.41
T2 (Malation 50 mg/kg + Vit C 200 mg/kg)	2337.043±44.83	73.02±4.16
T3(Vit C 200 mg/kg)	4720.748±267.42	54.63±4.80

DISCUSSION

In the present study, we evaluated MDA and glutathione levels in ovarian tissues of female rats by inducing malathion poisoning and assessed the protective effects of vitamin C. Based on the findings, experimental group 1, which only received malathion, experienced a significant increase in the level of MDA, compared to the control group ($P < 0.001$). These alterations can be explained by malathion-induced changes in the antioxidant system of cells and cell membrane lipid peroxidation, caused by the production of free radicals (4). Based on previous studies, organophosphates can cause changes in the biochemistry and histopathology of various organs (19, 20). Ovary is one of the target organs, which plays an important role in reproductive function through producing oocytes and hormones (21). The present findings were consistent with a similar study, which assessed the effects of malathion and endosulfan on lipid peroxidation of rat ovaries. It can be suggested that these toxins cause a significant increase in the level of MDA, confirming the production of free radicals and lipid peroxidation (22, 23). According to several previous studies, the increased level of MDA, induced by organophosphate intoxication, occurs in various tissues (e.g., heart, lung, liver, and brain tissues), and is accompanied by an increase in the activities of antioxidant systems in these tissues (24, 25). In addition, in the present study, the level of MDA increased in ovarian tissues of rats, treated with malathion, due to the production of free radicals and lipid metabolism (26). In the current study, comparison between experimental group 1 and the control group showed that the mean glutathione level was lower in experimental group 1 ($P < 0.001$). This difference seemed to be statistically significant. Previous studies have also confirmed the reduced amount of glutathione and increased MDA level in erythrocytes of mice and rats in the presence of fenthion, an organophosphate insecticide (25, 27). In our previous study on diazinon is an organophosphate poison was found that glutathione level significantly decreased and MDA level significantly increased in ovarian

tissue of female rats (28). Also, it has been shown that glutathione level is significantly decreased in the blood of malathion-treated rats (29). Moreover, a significant decline has been reported in glutathione level in blood and liver tissues due to malathion toxicity (30). Due to exposure to malathion, which is an organophosphate compound, the antioxidant system of the body becomes activated against ROS (31). Vitamin C is a water-soluble vitamin, which can reduce the amount of free radicals via its antioxidant features (32). In the present study, as previously stated, vitamin was intraperitoneally injected for 14 days in rats. In experimental group 3 (malathion and vitamin C), a significant increase in the level of glutathione and a significant decline in MDA level were reported, compared to the malathion group ($P < 0.002$). Therefore, we can conclude that vitamin C was able to improve oxidative stress. Based on the literature, use of vitamin E and C in rats, exposed to diazinon (which is an organophosphate toxin), can significantly improve lipid peroxidation in erythrocytes (33). Also, oxidative stress, induced by diazinon in brain tissues, is significantly decreased by vitamin C and E (34). In addition, it has been shown that vitamin C can prevent hepatotoxicity induced by methyl parathion in rats (35). Based on the findings, we can conclude that oxidative stress in malathion-treated rats can cause cellular toxicity. The need for treatment and compliance with the instructions is strongly felt for preventing the entry of toxins to the body, deterring toxicity-induced tissue damages, and preventing other disorders such as gonadal dysfunction and risk of sterility.

ACKNOWLEDGMENTS

The present study was extracted from an M.Sc. thesis of anatomical sciences, approved by Mashhad University of Medical Sciences (code: 922842). We would like to thank the Research Deputy of Mashhad University of Medical Sciences and Ms. Motejaded, the technician of histochemistry lab, for their sincere assistance and technical services. We also extend our

gratitude to the Department of Toxicology at the School of Pharmacy.

REFERENCE

- Hoffmann U, Papendorf T. Organophosphate poisonings with parathion and dimethoate. *Intensive care medicine*. 2006;32(3):464-68.
- Storm JE, Rozman KK, Doull J. Occupational exposure limits for 30 organophosphate pesticides based on inhibition of red blood cell acetylcholinesterase. *Toxicology*. 2000;150(1):1-29.
- Abdollahi M, Mostafalou S, Pournourmohammadi S, Shadnia S. Oxidative stress and cholinesterase inhibition in saliva and plasma of rats following subchronic exposure to malathion. *Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology*. 2004;137(1):29-34.
- Michiels C, Raes M, Toussaint O, Remacle J. Importance of Se-glutathione peroxidase, catalase, and Cu/Zn-SOD for cell survival against oxidative stress. *Free radical Biology and medicine*. 1994;17(3):235-48.
- Giordano G, Afsharnejad Z, Guizzetti M, Vitalone A, Kavanagh TJ, Costa LG. Organophosphorus insecticides chlorpyrifos and diazinon and oxidative stress in *Toxicology and applied pharmacology*. 2007;219(2):181-89.
- Kalender Y, Kaya S, Durak D, Uzun FG, Demir F. Protective effects of catechin and quercetin on antioxidant status, lipid peroxidation and testis-histoarchitecture induced by chlorpyrifos in male rats. *Environmental toxicology and pharmacology*. 2012;33(2):141-48.
- Behrman HR, Kodaman PH, Preston SL, Gao S. Oxidative stress and the ovary. *Journal of the Society for Gynecologic Investigation*. 2001;8(1 suppl):S40-S42.
- Possamai F, Fortunato J, Feier G, Agostinho F, Quevedo J, Wilhelm Filho D, et al. Oxidative stress after acute and sub-chronic malathion intoxication in Wistar rats. *Environmental toxicology and pharmacology*. 2007;23(2):198-204.
- Sabatini L, Wilson C, Lower A, Al-Shawaf T, Grudzinskas JG. Superoxide dismutase activity in human follicular fluid after controlled ovarian hyperstimulation in women undergoing in vitro fertilization. *Fertility and sterility*. 1999;72(6):1027-34.
- Ishikawa M. [Oxygen radicals-superoxide dismutase system and reproduction medicine]. *Nihon SankaFujinkaGakkaiZasshi*. 1993;45(8):842-48.
- Ogutcu A, Uzunhisarcikli M, Kalender S, Durak D, Bayrakdar F, Kalender Y. The effects of organophosphate insecticide diazinon on malondialdehyde levels and myocardial cells in rat heart tissue and protective role of vitamin E. *Pesticide biochemistry and physiology*. 2006;86(2):93-98.
- Hariri AT, Moallem SA, Mahmoudi M, Memar B, Hosseinzadeh H. Sub-acute effects of diazinon on biochemical indices and specific biomarkers in rats: protective effects of crocin and safranal. *Food and chemical toxicology*. 2010;48(10):2803-08.
- Couto N, Malys N, Gaskell SJ, Barber J. Partition and turnover of glutathione reductase from *Saccharomyces cerevisiae*: a proteomic approach. *Journal of proteome research*. 2013;12(6):2885-94.
- Sargazi Z, Nikravesh MR, Jalali M, Sadeghnia H, RahimiAnbarkeh F, Mohammadzadeh L. Gender-Related Differences in Sensitivity to Diazinon in Gonads of Adult Rats and the Protective Effect of Vitamin E. *International Journal of Women's Health and Reproduction Sciences*. 2015; 3 (1):40-47.
- Salehi B, Vakilian K, Ranjbar A. Relationship of Schizophrenia with Lipid Peroxidation, Total Serum Antioxidant Capacity and Thiol Groups. *Iranian Journal of Psychiatry and Clinical Psychology*. 2008;14(2): 5-14.
- Hubel CA. Oxidative stress in the pathogenesis of preeclampsia. *Experimental Biology and Medicine*. 1999; 222(3):222-35.
- Bilodeau J, Hubel C. Current concepts in the use of antioxidants for the treatment of

- preeclampsia. *Journal of obstetrics and gynaecology Canada: JOGC= Journal d'obstetriqueetgynecologie du Canada: JOGC.* 2003;25(9):742-50.
18. Ramos R, Gomez-Gerique N, Martinez-Castelao A. Lipoprotein oxidation profile in end stage renal disease patients. Role of vitamin C supplementation. *Nefrologia.* 2005;25(2):178-84.
 19. Gomes J, Dawodu A, Lloyd O, Revitt D, Anilal S. Hepatic injury and disturbed amino acid metabolism in mice following prolonged exposure to organophosphorus pesticides. *Human & experimental toxicology.* 1999;18(1):33-37.
 20. Yavuz T, Delibas N, Yildirim B, Altuntas I, Candir O, Cora A, et al. Vascular wall damage in rats induced by organophosphorus insecticide methidathion. *Toxicology letters.* 2005;155(1):59-64.
 21. Mattison DR, Thomford PJ. The mechanisms of action of reproductive toxicants. *Toxicologic pathology.* 1989;17(2):364-76.
 22. Fulia A, Chauhan P, Sharma R. Ameliorating effect of vitamin e on testicular toxicity induced by endosulphan in *Capra hircus* in vitro. *J PharmacoToxicol.* 2011;6(2):133-40.
 23. Koc N, Kayhan F, Sesal C, Muşlu M. Dose-dependent effects of endosulfan and malathion albino rat ovaries. *Pakistan journal of biological sciences: PJBS.* 2009;12(6):498-503. on adult wistar
 24. Buyukokuroglu ME, Cemek M, Yurumez Y, Yavuz Y, Aslan A. Antioxidative role of melatonin in organophosphate toxicity in rats. *Cell biology and toxicology.* 2008;24(2):151-58.
 25. Lukaszewicz-Hussain A. Role of oxidative stress in organophosphate insecticide toxicity—Short review. *Pesticide biochemistry and physiology.* 2010;98(2):145-50.
 26. Jahromi VH, Koushkaki M, Kargar H. The effects of malathion insecticide on ovary in female rats. *National park forschung in der schweiz (Switzerland Research Park Journal).* 2012;101(5):231-35.
 27. Yurumez Y, Cemek M, Yavuz Y, Birdane YO, Buyukokuroglu ME. Beneficial effect of N-acetylcysteine against organophosphate toxicity in mice. *Biological and Pharmaceutical Bulletin.* 2007;30(3):490-94.
 28. Sargazi Z, Nikravesh MR, Jalali M, Sadeghnia H, Anbarkeh FR, Mohammadzadeh L. Diazinon-Induced Ovarian Toxicity and Protection by Vitamins E. 2014; 8(26):1130-35.
 29. Ahmed RS, Seth V, Pasha S, Banerjee B. Influence of dietary ginger (*ZingiberofficinalesRosc*) on oxidative stress induced by malathion in rats. *Food and Chemical Toxicology.* 2000;38(5):443-45.
 30. Hazarika A, Sarkar S, Hajare S, Kataria M, Malik J. Influence of malathion pretreatment on the toxicity of anilofos in male rats: a biochemical interaction study. *Toxicology.* 2003;185(1):1-8.
 31. El-Demerdash FM. Lipid peroxidation, oxidative stress and acetylcholinesterase in rat brain exposed to organophosphate and pyrethroid insecticides. *Food and Chemical Toxicology.* 2011;49(6):1346-52.
 32. Sutcu R, Altuntas I, Yildirim B, Karahan N, Demirin H, Delibas N. The effects of subchronic methidathion toxicity on rat liver: role of antioxidant vitamins C and E. *Cell biology and toxicology.* 2006;22(3):221-27.
 33. sutcu R, Altuntas I, Buyukvanli B, Akturk O, Ozturk O, Koylu H, et al. The effects of diazinon on lipid peroxidation and antioxidant enzymes in rat erythrocytes: role of vitamins E and C. *Toxicology and industrial health.* 2007;23(1):13-17.
 34. Yilmaz N, Yilmaz M, Altuntas I. Diazinon-induced brain toxicity and protection by vitamins E plus C. *Toxicology and industrial health.* 2012;28(1):51-57.
 35. Kalender Y, Uzunhisarcikli M, Ogutcu A, Acikgoz F, Kalender S. Effects of diazinon on pseudocholinesterase activity and haematological indices in rats: the protective role of vitamin E. *Environmental toxicology and pharmacology.* 2006;22(1):46-51.